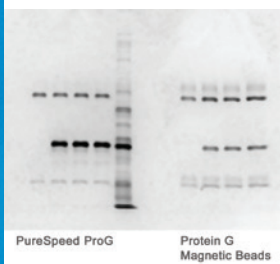


Master Your Workflow

High Performance IP with PureSpeed



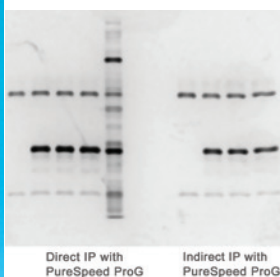
High quality data

High performance immunoprecipitation (IP) with PureSpeed and the E4 XLS pipette yields more concentrated immunoprecipitated protein than other techniques such as magnetic or agarose beads. Higher concentration increases confidence that the procedure is effective and reduces the likelihood of needing to repeat an experiment.



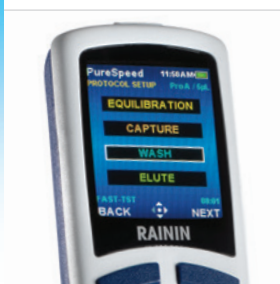
High throughput

Using the E4 XLS multichannel pipette and PureSpeed tips, immunoprecipitation protocol steps can be carried out in parallel rather than rapid succession. The PureSpeed IP system can carry out IP of 12 samples in less than 30 minutes with an optimized protocol.



Flexible protocols

Unlike many alternative technologies, Rainin has validated IP protocols for capturing pre-formed antigen-antibody complexes from lysates (indirect IP) as well as protocols for using preimmobilized antibody on ProA or ProG resin to capture antigen from lysates (direct IP).



Semi-automated system

Many current IP protocols require extensive pipetting, especially during wash steps. Semi-automation with the E4 XLS electronic pipette and PureSpeed tips decreases manual labor without the need for investing in high cost automation platforms.



PureSpeed™ IP System

Speed, Scalability, Dependability

High Performance IP with PureSpeed ProA and ProG tips and the E4 XLS electronic pipette can be used for both direct and indirect IP procedures. Compared with other IP technologies, the PureSpeed IP system offers greater protocol flexibility, workflow speed, and data quality while providing the benefits of semi-automation using the cost effective E4 XLS. The small footprint and light weight of the system allows it to be moved around the laboratory with ease.

PureSpeed IP with E4 XLS

- Easy customization of IP protocols using PureSpeed
- ProA or ProG tips to support different antibody subclasses
- Resin is immobilized within tips – no possible loss of resin during pipetting steps, unlike bead techniques
- Operable at room temperature and at 4 °C
- High levels of antigen-antibody complex can be isolated
- 96-deepwell plate for rapid screening of IP buffer conditions
- Low elution volumes: 15 – 240 µL depending on resin size
- Protocols can be transferred to other researchers.

Ordering Information

Catalog No.	MT Order No.	Description
PureSpeed Tips and Accessories		
PT-2-A5	17012561	ProA 5 µL Resin, 200 µL tip, 12 tips
PT-2-A20	17012562	ProA 20 µL Resin, 200 µL tip, 12 tips
PT-2-G5	17012563	ProG 5 µL Resin, 200 µL tip, 12 tips
PT-2-G20	17012564	ProG 20 µL Resin, 200 µL tip, 12 tips
PT-10-A20	17012568	ProA 20 µL Resin, 1000 µL tip, 12 tips
PT-10-A80	17012569	ProA 80 µL Resin, 1000 µL tip, 12 tips
PT-10-G20	17012570	ProG 20 µL Resin, 1000 µL tip, 12 tips
PT-10-G80	17012571	ProG 80 µL Resin, 1000 µL tip, 12 tips
PT-ACC	17012588	PureSpeed Accessories
LR-P2-96P-5	17012623	2.2 mL 96-Deepwell Plate
PureSpeed Single Channel Starter Kits		
PT-S2-A20	17012577	E4-200XLS + ProA PureSpeed Tips
PT-S2-G20	17012578	E4-200XLS + ProG PureSpeed Tips
PT-S10-A20	17012579	E4-1000XLS + ProA PureSpeed Tips
PT-S10-G20	17012580	E4-1000XLS + ProG PureSpeed Tips
PureSpeed Multichannel Starter Kits		
PT-S2-E8	17013548	E8-200 XLS and Accessory Kit
PT-S10-E8	17013546	E8-1200 XLS and Accessory Kit
PT-S2-E12	17013549	E12-200 XLS and Accessory Kit
PT-S10-E12	17013547	E12-1200 XLS and Accessory Kit

Technical Specifications

Tip sizes	200 µL	1000 µL
Resin bed volumes (µL)	5 or 20	20 or 80
Storage temperature	4°C	

Rainin PureSpeed tips fit E4 XLS pipettes with LTS.

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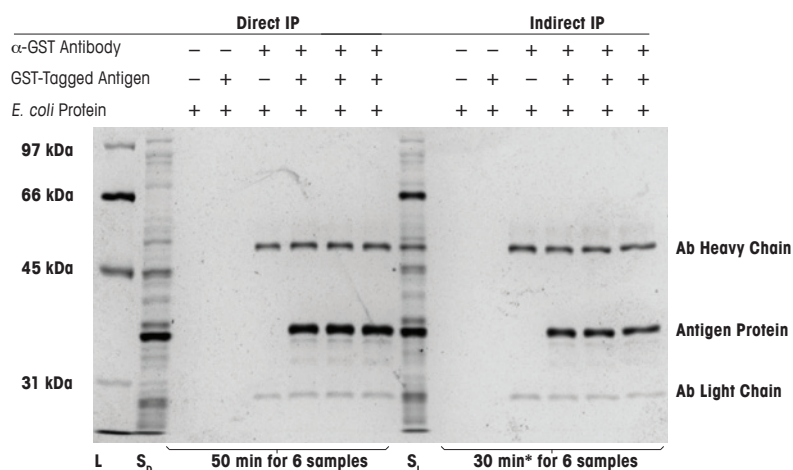
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Subject to technical changes

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Comparison of Direct and Indirect IP using PureSpeed.

In this experiment, 10 µg of antibody was used to immunoprecipitate 5 µg of GST-tagged antigen using the direct and indirect methods. The IP reactions occurred in the context of 125 µg of *E. coli* protein (total volume was 200 µL). The protein ladder is denoted as L, while the protein solutions (containing antigen for the direct method, and antigen and antibody for the indirect method) are labeled S₀ and S₁ (the D and I subscripts indicate the protein solutions for the direct and indirect methods). Sample eluates for direct and indirect IP are in lanes 3-8, and 10-15, respectively. Plus and minus indicate whether a certain component was included, or excluded in a given IP reaction. Lastly, the time at the bottom of the gel indicates the time needed for each protocol.

*The time for indirect IP does not include the 4 °C overnight incubation of IP samples.

www.mt.com/purespeed

For more information