BioClean

RAININ
Pipetting 360°

CERTIFICATE
OF QUALITY

Rainin BioClean™ pipette tips labeled "Certified RNase-, DNAse-, Endotoxin- and ATP-free" have been process-tested and passed the following detection levels using the test protocols below.

Contaminants Tested	Testing Detection Levels
RNase	≤ 10^{-9} Kunitz units/µL
DNAse	≤ 10^{-7} Kunitz units/µL
Human DNA	< 0.32 pg

Contaminants Tested	Testing Detection Levels
Bacterial DNA	< 1 pg
Endotoxin	< 0.001 EU/ml
ATP	< 2 X 10^{-12} mg/µl

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Head of Quality Systems

Testing Procedures

RNase testing protocol
Products were rinsed in DNase-, RNase-free 0.1 µm filtered distilled water, then product extracts were exposed to an RNA standard in a fixed volume of buffer. The RNA standard was incubated at 37°C for 1 hour. RNA fluorescence was measured using an RNA fluorescent dye and evaluated for degradation.

DNase testing protocol
Products were rinsed in DNase-, RNase-free 0.1 µm filtered distilled water, then product extracts were exposed to an DNA standard in a fixed volume of buffer. The DNA standard was incubated at 37°C for 1 hour. DNA fluorescence was measured using an DNA fluorescent dye and evaluated for degradation.

DNA testing protocol
Quantitative PCR (qPCR) was used in the following fashion: A BioRad CFX96 system was used to detect amplification in 20 µL reaction volumes containing negative controls, positive controls, varying concentrations of stock DNA (human or bacterial) and tip eluate. Final primer concentration is 200-300 nM. Both human and bacterial primer sets for conserved sequences were used, and are as follows:

Human primers: Forward: 5′-TGAATGGGAGAAGGCAGAAG
Reverse: 5′-TATCCACCGGGTGTTTTCTC

Bacterial primers: Forward: 5′-CAAGGCTAAATACTCCTGAC
Reverse: 5′-CACTCCCCTCGCCGGGGTTC

Endotoxin testing protocol
Products are extracted in endotoxin-free LAL reagent water for 1 hour, then product extracts are tested by kinetic assay. The test is performed by adding LAL to the negative control, Control Standard Endotoxin, positive control and product extracts. After a fixed incubation period, the reaction mixture is measured. The sensitivity of the kinetic assay is 0.001 EU/mL.

ATP tested by the following protocol
ATP is tested by measuring the difference between baseline and product-rinsed luminescence of an ATP standard solution containing 10^{-11} mg of ATP. Perturbations in light emission of a product-rinsed ATP standard solution are evaluated to determine the presence or absence of ATP.

Sterilized product
Rainin tip products labeled as "sterilized" are irradiated by gamma radiation (SAL=10^{-6}). Dosage level has been predetermined by bioburden testing.

Limitation of Liability
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